# The Palaearctic triozids associated with Rubiaceae (Hemiptera, Psylloidea): a taxonomic re-evaluation of the Trioza galii Foerster complex 

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The Palaearctic triozids associated with Rubiaceae (Hemiptera, Psylloidea): a taxonomic re-evaluation of the Trioza galii Foerster complex. - The Palaearctic Trioza galii complex is revised to contain T. cocquempoti sp. n., T. drosopouli sp. n., T. galii Foerster and T. velutina Foerster stat. rev., with following new synonymies: T. rubiae Baeva and T. rubicunda Loginova $=$ T. galii and Trioza distincta Flor $=$ T. velutina, respectively. T. galii f. spinogalii Sulc and T. galii f. aspinovelutina Šulc are considered to be of infrasubspecific rank and thus unavailable. Trioza velutina var. thoracica Flor is an available name regarded as nomen dubium. Lectotypes are designated for Trioza galii and T. velutina. Adults and last instar larvae (except T. velutina) are diagnosed and illustrated. Identification keys are provided. All four species are associated with Rubiaceae on which they induce galls. Confirmed hosts of T. galii are Galium spp. and Asperula cynanchica, literature records also suggest Sherardia arvensis and Rubia spp. T. cocquempoti and T. drosopouli develop on Rubia spp. T. velutina is probably associated with Galium spp. but hard evidence is not available. The T. galii complex is diagnosed using adult and larval charcters. The definition is similar to Conci's (1992) concept of Spanioza but excludes S. tamaninii. S. tamaninii is morphologically intermediate between the T. galii complex and the T. centranthi complex (associated with Valerianaceae) which may be sister groups. The synonymy of Spanioza with Trioza is confirmed, and the new combination Trioza tamaninii is proposed.
Keywords: Hemiptera - Psylloidea -Triozidae - taxonomy - new taxa Rubiaceae - Palaearctic.

## INTRODUCTION

Species of jumping plant-lice are generally well-defined by their adult and larval morphology as well as their host plants. In particular, the male terminalia are routinely used for species identification. The Palaearctic Trioza galii Foerster, as
currently perceived, is a notable exception as the male genital morphology exceeds the usual variability encountered in other Trioza species.

Foerster (1848) described both T. galii and T. velutina from German and Irish localities; some material of $T$. galii was collected on Galium verum. The descriptions contain colour and forewing characters but are not diagnostic. Hardy (1853) recorded deformations on Galium aparine which he attributed to "Psylla" velutina. He said that velutina appears not to differ from "Psylla" galii. Flor (1861a), providing detailed descriptions and diagnoses, treated T. galii and T. velutina as distinct species, pointing out differences in the male paramere. He also assigned a single female specimen from Southern France with a lighter thorax and clear forewing colour, to the variety Trioza velutina var. thoracica. Again on the basis of a single female, he described Trioza distincta from Thuringia, differing from T. velutina in the slightly different forewing shape (Flor, 1861b). Flor's more restricted species concept was confirmed by Löw (1882) who examined some of Foerster's and Flor's types. Lethierry (1874), Oshanin (1907) and Aulmann (1913) followed Löw, whereas Scott (1876) and Edwards (1896) adopted Hardy's view, treating T. galii and velutina as synonyms. Šulc (1913) pointed out that his earlier description (Šulc, 1910) of T. galii corresponded with the types of T. velutina and having examined types of T. galii, T. velutina and T. distincta, plus other material, reached the same conclusion as Hardy (1853). Thus, he treated T. velutina, T. distincta and questionably var. thoracica, the type of which appears to be lost, as synonyms of T. galii. Based on presence or absence of surface spinules as well as colour and shape of the forewing, he defined four forms: galii forma typica, f. spinogalii Šulc, f. velutina and f. aspinovelutina Šulc. Boselli (1930) found many specimens in Italy on Rubia peregrina which he referred to Spanioza galii aspinovelutina. He gave detailed descriptions and illustrations of the adults, larvae and eggs and provided information on their biology. The male paramere of Boselli's material, however, is more massive than those described and illustrated by Šulc (1910, 1913). T. galii, in this broad definition, has been reported from all over the Palaearctic, including Japan and Taiwan (Aulmann, 1913; Klimaszewski, 1973; Gegechkori \& Loginova, 1990; Ossiannilsson, 1992). According to these authors, the species is associated with Galium, Sherardia and Rubia spp. (Rubiaceae), where it induces characteristic deformations on the buds, shoots and leaves, which can be above or below ground (e.g. Houard, 1909; Docters van Leeuwen, 1937).

Adults are regularly encountered by sweeping herbaceous vegetation, although rarely on the host plants. Larval material, by contrast, is generally difficult to find. The first larval description referred to T. galii is by Kieffer (1889) who examined material from Sherardia arvensis but his description is not diagnostic. The dorsum of the larva is said to be dark brown. A more detailed larval description referred to T. galii is that by Boselli (1930). White \& Hodkinson (1981) used Boselli's description in their handbook of the British species, and Ossiannilsson (1992) in the Fauna Scandinavica described Italian rather than Scandinavian specimens.

Three additional Palaearctic species are reported from Rubiaceae. Baeva (1972) described Trioza rubiae from Turkmenia based on a single series collected on Rubia florida, which she related to Trioza foersteri Meyer-Dür and Trioza rotundata Flor. Loginova (1978) added Trioza rubicunda, again based on a single series, collected in

Kazakhstan on Galium sp. According to Loginova T. rubicunda is closest to T. galii f. velutina from which it differs in the reduced surface spinules and a broader paramere. T. rubiae and T. rubicunda have not been subsequently reported. Conci (1992) described Spanioza tamaninii from Italian and Slovakian specimens, one of which was collected on Galium anisophyllon. He suggested that S. tamaninii is closely related to galii and that its host may be Galium.

Trioza galii was selected by Enderlein (1926) as type species of Spanioza, a genus which he erected for triozids with a short Rs vein in the forewing. The artificial nature of this concept was pointed out by Tuthill (1943) who synonymised it with Trioza. Conci (1992) considered Spanioza a valid genus and redefined it to include Trioza galii and its "formae", Trioza rubiae Baeva, Trioza rubicunda Loginova and Spanioza tamaninii Conci. This was not followed by Burckhardt \& Couturier (1994) who treated the Trioza galii group as part of the large artificial genus Trioza.

Over the last few years collections from the Mediterranean as well as Central and Western Europe yielded material including larvae and host records which permit a re-evaluation of the taxonomy of the Trioza galii complex. Wagner \& Franz (1961) suggested that T. galii in the broad definition by Šulc (1913) is a species complex, and Conci (1992) wrote that Trioza galii and its "formae" require revision. The present paper provides this revision and re-examines the validity of Spanioza.

## MATERIAL AND METHODS

Material has been examined or is mentioned from following institutions:
BMNH Natural History Museum, London
DEZA Dipartimento di Entomologia e Zoologia Agraria dell'Università, Portici
EAUT Institute of Zoology and Botany, Estonian Agricultural University, Tartu
MHNG Muséum d'histoire naturelle, Geneva
MMBC Moravské Muzeum, Brno
MNHN Muséum national d'histoire naturelle, Paris
NHMB Naturhistorisches Museum, Basel
NHMV Naturhistorisches Museum, Vienna
SMTD Staatliches Museum für Tierkunde, Dresden
ZISP Zoological Institute, St. Petersburg
ZMHB Museum für Naturkunde der Humboldt-Universität, Berlin
ZSSM Zoologische Staatssammlung, Munich.
The morphological terminology mainly follows Ossiannilsson (1992). Drawings were prepared from cleared specimens mounted permanently in Euparal or Canada Balsam or temporarily in glycerine. Measurements were taken from slide mounted specimens, except for the total body length which is taken from dry mounted specimens measured from the apex of head to the apex of forewing when folded over the body.

## RESULTS

The Trioza galii Foerster, 1848 species complex

## DESCRIPTION

Adult. Coloration mostly dark brown to black, in T. cocquempoti and T. drosopouli thorax yellowish, brown or dark dorsally, sometimes with longitudinal
stripes; area around wing insertion yellowish or brown; intersegmental membranes reddish; antennal segment 3 , tibiae and tarsi whitish. Forewing almost colourless or with yellowish or light brown tinge; veins yellowish or light brown, vein $\mathrm{C}+\mathrm{Sc}$ often brown or dark brown in basal part. Young specimens lighter.

Head slightly narrower than mesonotum, weakly inclined from longitudinal body axis, at a $45^{\circ}$ angle. Vertex shiny, weakly produced anteriorly on either side of mid-line. Genal processes variable in size and shape, conical, subacute, ranging from one to two thirds vertex length. Antenna 10 -segmented, segment 3 longer than segments 4 and 5 together; segment 10 bearing one long and one very short terminal seta respectively (Fig. 21). Clypeus short, pyriform. Thorax weakly curved dorsally. Metacoxa with horn-shaped, subacute meracanthus; metatibia weakly widened basally and apically, with some minute basal spines and $1+3$ sclerotised apical spurs; metabasitarsus without sclerotised spurs. Forewing (Figs 1-4) transparent, variable in size, shape and colour, elongate, strongly pointed, angular or bluntly angular apically; vein Rs short, bifurcation of vein M slightly or strongly distal to line linking apices of veins Rs and $\mathrm{Cu}_{1 \mathrm{a}}$. Wing membrane transparent; apart from wing base usually lacking surface spinules, with the exception of T. velutina where spinules are always present, forming more or less extended fields. Radular spinules forming narrow stripes in cells $\mathrm{m}_{1}, \mathrm{~m}_{2}$, and $\mathrm{cu}_{1}$. Hindwing membranous, about two thirds forewing length. Costal setae ungrouped; $\mathrm{R}+\mathrm{M}+\mathrm{Cu}_{1}$ indistinctly trifurcating. Abdominal tergite 3 (and occasionally 4 ) in male and tergite 4 (and occasionally 5 ) in female bearing a row of lateral setae. Male proctiger (Figs 5, 8, 11, 16) 1-segmented, weakly produced or lobed posteriorly, covered in moderately long setae apically and posteriorly. Subgenital plate subglobular, with a few long setae laterally and ventrally. Paramere (Figs 6, 9, 12, 13, 17) shorter than proctiger, massive or lamellar; sparsely covered in long setae on outer face apically and along hind margin; inner face covered in long thin setae mainly basally, apically and along hind margin, with a group of shorter, stout setae along fore margin in basal half. Proximal portion of aedeagus strongly curved at base, almost straight apically; distal portion (Figs 7, 10, 14, 15, 18) relatively short with large apical dilatation which varies from almost rectangular to distinctly hooked. Sclerotised end tube of ductus ejaculatorius short, weakly curved. Female proctiger (Figs 23-26) short with relatively large circumanal ring. Subgenital plate short.

Fifth instar larva (Fig. 28). Coloration varying from uniformly yellow or light ochreous to yellow with more or less extended pattern on head, thorax, wing buds and abdomen to completely dark. Body oval, bearing a single row of densely spaced, lateral truncate sectasetae. Antenna 6 -segmented. Head broadly rounded anteriorly. Legs with short claws and small, fan-shaped tarsal arolium. Humeral lobes blunt, relatively short. Circumanal ring small, with a single row of pores. Dorsum of body lacking macroscopic setosity. Tarsal arolium transversely oval, lacking pedicel, with unguitractor (Figs 29-31). Claws relatively small.

Egg. Spindle-shaped, longer than three times its width, with short basal pedicel and no apical filament. The egg is laid perpendicularly on the leaf margins.


Figs 1-4
Forewing of Trioza spp. 1-T. cocquempoti; 2 - T. drosopouli; $3-$ T. galii; $4-$ T. velutina.

## DISCUSSION

The T. galii complex, as defined above, constitutes a small, probably monophyletic group within the large artificial genus Trioza. It is restricted to the Palaearctic. It may be most closely related to the $T$. centranthi complex ( $T$. centranthi (Vallot), Trioza nana Gegechkori and T. valerianae Gegechkori), associated with Valerianaceae. Adult T. galii and T. velutina resemble T. centranthi (Vallot) in morphology, size and colour. The distribution of surface spinules on the forewing is variable in T. centranthi. They may form relatively extended fields in all cells except for $\mathrm{c}+\mathrm{sc}$ or they may be restricted to cell $\mathrm{cu}_{2}$. In $T$. centranthi vein Rs of the forewing tends to be relatively long and sinuous, and the bifurcation of vein M lies near the line joining the apices of veins Rs and $\mathrm{Cu}_{1 \mathrm{a}}$; in T. galii and T. velutina in contrast vein Rs is shorter and almost straight or concave; the bifurcation of vein M is more distant to the line joining the apices of veins Rs and $\mathrm{Cu}_{1 \mathrm{a}}$. T. centranthi bears terminal setae on antennal segment 10 , with the shorter seta about half as long as the longer one (Fig. 22), in the T. galii complex the shorter seta is only about a quarter to a third as long as the longer (Fig. 21). The T. centranthi complex is characterised by the irregularly triangular paramere, which sometimes bears a large antero-apical lobe (absent in the T. galii complex), and by the apical dilatation of the distal portion of the aedeagus, which bears small lateral spines ( $T$. centranthi, Fig. 20) (absent in the T. galii complex, Figs 7, 10, 14, 15, 18). The female subgenital plate of T. centranthi (Fig. 27) has, on either side, a group of long setae near the dorsal margin, which are longer than the other hairs (hairs laterally, ventrally and terminally are of about the same length in the T. galii complex, Figs 23-26).

The present concept of the T. galii complex differs in two major details from that of Conci (1992). In addition to T. galii, T. rubiae and T. rubicunda, Conci included Spanioza tamaninii in the group. S. tamaninii has so far been recorded from a few specimens from Italy and Slovakia only. It differs from the T. galii complex and the T. centranthi complex in having antennae which have light segments 3-8 rather than dark segments with a strongly contrasting light segment 3 . The paramere shape, the termi-


Figs 5-10
Male genital structures of Trioza spp. 5-7 - T. cocquempoti; 8-10 - T. drosopouli. 5, 8 Terminalia, in profile, scale a; 6, 9 - paramere, inner surface, scale b; 7, 10 - distal portion of aedeagus, scale b.
nal antennal setae and probably also the lateral setosity on the female subgenital plate are as in species of the $T$. centranthi complex. Apart from the antennal colour $S$. tamaninii differs from the T. centranthi complex in the apex of the aedeagus (Fig. 19) lacking lateral spines (cf. Fig. 20 for T. centranthi). Incidentally Conci's drawing of the aedeagus appears to be artefactual. Larval material is necessary to determine the phylogenetic relationship of S. tamaninii.


Figs 11-20
Male genital structures of Trioza spp. 11-15 - T. galii; 16-18 - T. velutina; 19-T. tamaninii; 20 -T. centranthi. 11, 16 - Terminalia, in profile, scale a; 12, 13, 17 - paramere, inner surface, scale b; 14, 15, 18-20 - distal portion of aedeagus, scale b.

Conci (1992) resurrected the genus Spanioza for the T. galii complex and S. tamaninii. He neither provided convincing autapomorphies for Spanioza nor did he define a sister group. This makes his Spanioza highly artificial and without phylogenetic significance. We follow Burckhardt \& Couturier (1994) who treated Spanioza as a synonym of Trioza. Here we formally transfer Spanioza tamaninii to Trioza as $T$. tamaninii (Conci) comb. n.

## TAXONOMIC TREATMENT

Key for the identification of the species of the Trioza galii complex
Adults
1 Antenna longer than 1.1 mm . Male proctiger strongly produced posteriorly (Figs 5, 8). Paramere (Figs 6,9) massive, blunt apically. Apex of distal portion of aedeagus subrectangular or reniform (Figs 7, 10). Ventral and posterior margin of female subgenital plate, in profile (in cleared specimens), forming an angle $>90^{\circ}$ (Figs 23, 24)

- Antenna shorter than 1.1 mm . Male proctiger tubular, weakly produced posteriorly (Figs 11, 16). Paramere (Figs 12, 13, 17) slender, with apical digitiform process. Apex of distal portion of aedeagus hooked (Figs 14, $15,18)$. Ventral and posterior margin of female subgenital plate, in profile (in cleared specimens), forming an angle $<90^{\circ}$ (Figs 25, 26)3

2 Male proctiger with narrow posterior lobe (Fig. 5). Paramere (Fig. 6), in profile, with narrow angular bulge above the middle along the fore margin; apex angular. Distal portion of aedeagus (Fig. 7) with reniform apical dilatation. Female terminalia as in Fig. 23. Algeria, France, Italy, Morocco, Spain T. cocquempoti sp. n. Male proctiger with large posterior lobe (Fig. 8). Paramere (Fig. 9), in profile, with broad bulge in apical third along the fore margin; apex broadly rounded. Distal portion of aedeagus (Fig. 10) with subrectangular apical dilatation. Female terminalia as in Fig. 24. Greece
T. drosopouli sp. n.

3 Forewing (Fig. 3) membrane lacking surface spinules except for base of cell $\mathrm{cu}_{2}$. Paramere robust, fore margin irregularly curved in apical half (Figs 12, 13). Apex of distal portion of aedeagus with large apical hook (Figs 14, 15). Female proctiger with relatively long and slender apical process, subgenital plate, in profile, truncate apically (Fig. 25) T. galii Forewing (Fig. 4) membrane with surface spinules present in all cells forming more or less extended fields. Paramere slender, evenly curved (Fig. 17). Apex of distal portion of aedeagus with relatively short apical hook (Fig. 18). Female proctiger with relatively short massive apical process, subgenital plate, in profile, bearing a small point apically (Fig. 26)
T. velutina

Fifth instar larvae
(larvae of T. velutina unknown)
1 Body dimensions small, body length $<1.8 \mathrm{~mm}$, see also table $2 \ldots$. . . . T. galii

- Body dimensions large, body length $>1.8 \mathrm{~mm}$, see also table $2 \ldots . \ldots$. . . . . 2

2 Marginal sectasetae on forewing pads over 100, and on hindwing pads over 15 . West Mediterranean T. cocquempoti $\mathrm{sp} . \mathrm{n}$. Marginal sectasetae on forewing pads less than 100, and on hindwing pads less than 15 . Greece T. drosopouli sp. n.

## Trioza cocquempoti sp. n .

Figs 1, 5-7, 23, 29
Spanioza galii aspinovelutina sensu Boselli, 1930: 14, nec Šulc, 1913.
Trioza galii sensu White \& Hodkinson, 1982: 43, nec Foerster, 1848.
Material examined: Holotype ठ̄, France: Gard, 5 km W Pont-St.-Esprit, 22.x.1988, Galium sp. (D. Burckhardt) (MHNG, dry mounted).

Paratypes. Algeria: 1 f, Atlas of Blida Chréa, Les Glacières, 1100 m , 3.v. 1988 (D. Burckhardt) (MHNG, dry mounted); 1 \&, Grande Kabylie, Oued Isser, bridge N24, 21.v. 1988 (D. Burckhardt) (MHNG, dry mounted). - France: 2 ㅇ, Corsica, Luri, Col de Sainte Lucie, 24. iv. 1992 (W. Della Giustina) (NHMB, dry mounted); 1 \&, Gard, same data as holotype; 1 ō, 1 ¢, Var, Roquebrune-sur-Argens, Les Sauterons, 15-20.iv. 1990 (C. Lienhard) (NHMB, dry mounted); 1 すै, 2 ¢, Indre et Loire, La Roche-Clermaut, Malaise trap, 11.viii. 1989 (C.


Figs 21-27
Trioza spp. 21, 25 - T. galii; 22, 27- T. centranthi; 23 - T. cocquempoti; 24 - T. drosopouli; 26 - T. velutina. 21, 22 - Antennal segment 10, scale a; 23-27 - female terminalia, in profile, scale b.

Cocquempot) (MMBC, dry mounted and stored in glycerine); 1 ô, Indre et Loire, Roquebrunne-sur-Argens, 15-20.iv. 1990 (C. Cocquempot) (MMBC, stored in glycerine); 2 , Tarn, Albi, Fargues, 30.v.1933, Crataegus (A. Perrier) (MNHN, dry mounted); 1 \&, Tarn, Albi, 7.vi., Crataegus (A. Perrier) (MNHN, dry mounted); 1 ㅇ, Albi, Castelnau-de-Lévi, 8.iv., Rhus (A. Perrier) (MNHN, dry mounted); 1 ¢, Albi, Creyssence, 14.vi., Buxus (A. Perrier) (MNHN, dry mounted); 1 ठै, Vaucluse, La Roque-Alric, near Beaumes-de-Venise, 300 m , 23.iv. 1984 (C. Lienhard) (MHNG, slide mounted); 2 す', Vaucluse, near Vacqueyraz, river Ouvèze, 60 m , 26.iv. 1984 (C. Lienhard) (MHNG, dry mounted); 6 ठ̂, 7 ¢, without locality data (MNHN, dry mounted). - Italy: 1 ō, 5 ㅇ, 2 parasitised larvae, Portici, 25.v.1928, Rubia (F. B. Boselli) (DEZA, dry mounted); ca. 100 larvae and exuviae, 21.v.1928, Rubia (F. B. Boselli) (DEZA, alcohol); 10 む, 9 ㅇ, 45 larvae and exuviae, Portici, Parco Gussone, v-vi. 2004 (P. A. Pedata) (DEZA, alcohol; NHMB, slide mounted); 1 §, Basilicata, Potenza, PR Gallipoli Cognato, near Monte Croccia, 810-910 m, 1-4.v.2001, Malaise trap 2 (Springate et al.) (DEZA, alcohol); 1 §̂, Basilicata, Matera, PR Gravina di Matera, 15 km E of Masseria San Francesco, 16-19.iii.2002,

290-310 m, Malaise trap (Springate et al.) (DEZA, alcohol); 2 ㅇ, same but 27-30.iv.2001, 290$370 \mathrm{~m} ; 2$ रे, 2 q, same but PR Gravina di Matera, 2.5 km W of road to Ginosa, 28-30.iv.2001; 1 ¢, same but PR Gravina di Matera, 1.5 km W of road to Ginosa, 25-28.ix.2001; 1 on, Campania, Napoli, Parco Gussone, $55 \mathrm{~m}, 1-4 . \mathrm{iv} .2002$, Malaise trap (Springate et al.) (DEZA, MMBC, NHMB, alcohol, dry mounted); 2 §', 4 f, same but $55-80 \mathrm{~m}, 17-20 . \mathrm{v} .2001 ; 3$, 9 , same but Caserta, WWF San Silvestro, $270-300 \mathrm{~m}$, 13-16.v. 2001. - Morocco: 1 \& without wings, Tanger (MMBC, Šulc collection). - Spain: $1 \begin{gathered}\text { oै, Zaragoza Province, Monegros Region, Pina de }\end{gathered}$ Ebro, 360 m, 2.vi. 1990 (J. Blasco-Zumeta) (MHNG, dry mounted).

Material excluded from type series. France: 1 adult without abdomen, Vaucluse, La Roque-Alric, 350 m , 27.iv. 1984 (C. Lienhard) (MHNG, dry mounted).

Diagnosis: Adult. Total body length $2.8-3.3 \mathrm{~mm}$. Colour of thorax yellow, sometimes brown or dark brown dorsally, or with brown longitudinal stripes on mesopraescutum and mesoscutum. Genal processes 0.6 times as long as vertex along midline, subacute apically. Forewing (Fig. 1) elongate, pointed apically; bifurcation of vein M distinctly distal to line connecting apices of veins Rs and $\mathrm{Cu}_{1 \mathrm{a}}$. Forewing membrane lacking surface spinules except at wing base. Hindwing two thirds of forewing length. Male proctiger (Fig. 5) relatively massive, strongly produced posteriorly. Paramere (Fig. 6) massive, truncate apically, in profile, with narrow angular bulge in apical third along the fore margin; apex angular. Distal portion of aedeagus (Fig. 7) with reniform apical dilatation. Female proctiger (Fig. 23) relatively short, pointed apically; ventral and posterior margin of female subgenital plate, in profile (in cleared specimens), forming an angle $>90^{\circ}$. Measurements as in Table 1.

Fifth instar larva described by Boselli (1930) as Spanioza galii aspinovelutina. With following numbers of marginal sectasetae (one side only): head 42-46, forewing pad 109-113, hindwing pad 15-18, caudal plate 108-122. Tarsal arolium transversely oval (Fig. 29). Measurements as in Table 2.

Etymology: The species is dedicated to Christian Cocquempot, one of the collectors of the new species.

## Distribution: Algeria, France, Italy, Morocco, Spain.

Bıology. Host plant. Boselli (1930) collected eggs, larvae and adults exclusively on Rubia peregrina. The larvae induce a strong curling of the leaves. Boselli could not find any specimens on Galium. The holotype $\delta$ and a female paratype from France have been taken on Galium sp. but no larvae were found. At the moment there is no evidence that Galium is a host plant.

According to Boselli (1930) larvae of T. cocquempoti aestivate and hibernate in the galls on the host, which also has a summer and winter dormancy. In spring and autumn when the host produces new flush, the adults emerge and, a few days later after copulation, the females lay their eggs. The eggs are laid perpendicularly on the leaf margins or stems. Depending on the season the egg development takes two (spring generation) to over four weeks (autumn generation). If the conditions in spring are particularly favorable, three generations per year are possible.

Comments: The description of Spanioza galii aspinovelutina by Boselli (1930) concerns T. cocquempoti, which was also confirmed by the examination of Boselli's material. For the validity of the name aspinovelutina see comments to T. galii. The larval description by White \& Hodkinson (1982), tentatively referred to T. galii, is based on Boselli's (1930) description and, hence, refers to T. cocquempoti.


FIGS 28-31
Trioza spp. 28, 31 - T. galii; 29 - T. cocquempoti; 30 - T. drosopouli. 28 - Last instar larva, left dorsal face, right ventral face (scale a), with details of marginal sectasetae and tarsal apex (scale b); 29-31 - tarsal arolium, scale b.

Trioza drosopouli sp. n . Figs 2, 8-10, 24, 30
Material examined: Holotype đ', Greece: Attiki, Tatoi, NW Athens, $250 \mathrm{~m}, 1 . \mathrm{v} .1995$, Rubia tinctorum (P. Lauterer) (NHMB, dry mounted).

Paratypes. Greece: $16 \delta^{\star}, 43$ ¢, 1 exuvia, Attiki, same data as holotype (MMBC, 14 ठर, 38 아 dry mounted, 1 ठ, 2 ㅇ, 1 exuvia, stored in glycerine; NHMB, 2 ㅇㅇ dry mounted, $1 \delta^{\star}, 1$ ㅇ stored in glycerine); 4 ठ, 12 , same data but $200 \mathrm{~m}, 7 . \mathrm{v} .1995$, clearing and Pinus wood
 Attiki, Paríntha Mountains, Agía Trías Parínthos, $1200 \mathrm{~m}, 28 . \mathrm{v} .1995$, Rubia tinctorum in wood
 ㅇ, Akarnanía, Gavrolimni, $120 \mathrm{~m}, 17 . \mathrm{iii} 1982$ (C. Lienhard) (MHNG, dry mounted); 1 ㅇ, Arkadia, E Peloponissos, 4 km E of Agios Petros, 900 m , $5 . \mathrm{v} .1995$ (P. Lauterer) (MMBC, dry mounted); 1 exuvia, Messinía, Messene, between Kalamata and Pilos, 16.v. 1979 (D. Burckhardt) (MHNG, slide mounted).

Material not included in type series. Greece: $1 \delta, 11$, damaged, Attiki, same data as holotype (MMBC); 1 very teneral $\xlongequal{\circ}$, Messinía, Messene, between Kalamata and Pilos, 16.v. 1979 (D. Burckhardt) (MHNG, slide mounted).

Diagnosis: Adult. Total body length $2.8-3.4 \mathrm{~mm}$. Colour of thorax yellow or ochreous; mesopraescutum dark brown or black, somtimes with a median longitudinal lighter stripe; mesoscutum ochreous to light brown, sometimes with dark brown longitudinal stripes, generally lighter than mesopraescutum. Genal processes 0.8 times as long as vertex along mid-line, subacute apically. Forewing (Fig. 2) elongate, pointed apically; bifurcation of vein M distinctly distal to line connecting apices of veins Rs and $\mathrm{Cu}_{1 \mathrm{a}}$. Forewing membrane lacking surface spinules except at wing base. Hindwing two thirds of forewing length. Male proctiger (Fig. 8) massive, strongly produced posteriorly. Paramere (Fig. 9) massive, blunt apically, in profile, with large angular bulge in apical third along the fore margin; apex rounded. Distal portion of aedeagus (Fig. 10) with angular apical dilatation. Female proctiger (Fig. 24) relatively short, pointed apically; ventral and posterior margin of female subgenital plate, in profile (in cleared specimens), forming an angle $>90^{\circ}$. Measurements as in Table 1.

Fifth instar larva. With following numbers of marginal sectasetae (one side only): head 39-44, forewing pad 94-99, hindwing pad 11-12, caudal plate 97-110. Tarsal arolium oval (Fig. 30). Measurements as in Table 2.

Etymology: The species is dedicated to our friend Sakis Drosopoulos who organised the collecting trip of Pavel Lauterer to Greece and assisted in the field.

## Distribution: Greece (Akarnanía, Arkadia, Attiki, Messinía).

Biology: A larval skin and adults, including teneral specimens, have been collected on Rubia tinctorum which is a likely host. A few teneral adults were collected on 1 May and followed by mature adults up to 28 May, suggesting that the bionomics may be similar to that of T. cocquempoti.

Comments: No reliable characters could be found for separating the females of T. drosopouli and T. cocquempoti.

Trioza galii Foerster
Figs 3, 11-15, 21, 25, 28, 31
Trioza galii Foerster, 1848: 87; Šulc, 1913: 38, p. p. Lectotype ő, Germany: NordrheinWestfalen, near Aachen (A. Foerster) (MMBC), here designated, examined.
Psylla galii; Hardy, 1853: 3876.

Trioza galii Foerster f. aspinovelutina Šulc, 1913: 44. Name unavailable, articles 45.6.1 and 45.6.4, ICZN, 1999.

Spanioza galii; Enderlein, 1926: 400; Conci, 1992: 258.
Trioza rubiae Baeva, 1972: 63. Holotype ठ, Turkmenistan: north-western part of TurkmenKhorasan Mountains, around Yaradzha, 21.iv.1971, Rubia florida (V. Baeva, O. Rudenko) (ZISP, dry mounted), examined. Syn. n.

Trioza rubicunda Loginova, 1978: 111. Holotype ठ̄, Kazakhstan: Chimkent District, AksuDzhibagly Reservation, Talass Mountain Range, 80 km E Chimkent, 10.vi.1966, Galium sp., (M. M. Loginova) (ZISP, dry mounted), examined. Syn. n.
Spanioza rubiae; Conci, 1992: 258.
Spanioza rubicunda; Conci, 1992: 258.
Material examined: Material was examined from following countries: Algeria; Armenia; Austria: Oberösterreich, Steiermark; Bulgaria; Czech Republic, Bohemia, Moravia; Cyprus; France: Finistère, Haute-Garonne, Hautes-Pyrénées, Hérault, Indre et Loire, Landes, Maine-et-Loire, Nord, Pas-de-Calais, Tarn, Var; Germany: Baden-Württemberg, Berlin, Brandenburg, Hessen (including paralectotype ठ̄, Ems, leg. C.H.G. von Heyden), NordrheinWestfalen (including $\delta^{6}$ lectotype and paralectotypes 6 ot and 1 adult without abdomen, Aachen), Sachsen; Greece: Atikí, Corfu, Crete, Cyclades, Évia; Israel; Italy: Alto Adige, Puglia; Jordan; Kazakhstan (including of holotype of Trioza rubicunda); Kyrgyzstan; Malta; Slovakia; Slovenia; Spain: Caceres, Zaragoza, Canaries; Switzerland: Aargau, Basel-Land, Genève, Luzern, Neuchâtel, Vaud, Valais; Turkey: Aydin; Tunisia; Turkmenistan (including ơ holotype of Trioza rubiae); United Kingdom: England, Scotland, Wales (BMNH, MHNG, MMBC, MNHN, NHMB, NHMV, SMTD, ZISP, ZMHB, ZSSM).

Diagnosis: Adult. Total body length 2.3-3.2 mm. Colour of thorax usually dark brown or almost black with exception of area around wing insertion which is yellowish. Younger specimens with light colour more expanded and dark areas brown rather than black. Genal processes 0.3-0.7 times as long as vertex along mid-line, blunt to subacute apically. Forewing (Fig. 3) varying from relatively short and wide to elongate, angular apically; bifurcation of vein M on or slightly distal to line connecting apices of veins Rs and $\mathrm{Cu}_{1 \mathrm{a}}$. Forewing membrane lacking surface spinules except at wing base. Hindwing three quarters or four fifth of forewing length. Male proctiger (Fig. 11) relatively slender, weakly produced posteriorly. Paramere (Figs 12, 13) lamellar, with an antero-basal bulge and subparallel margins in the middle third, apical third abruptly narrowed to posteriorly directed apex. Distal portion of aedeagus (Figs 14,15 ) with large hook-shaped apical dilatation. Female proctiger (Fig. 25) with relatively short thick apical process; subgenital plate, in profile, bearing small point apically. Measurements as in Table 1.

Fifth instar larva (Fig. 28). With following numbers of marginal sectasetae (one side only): head 34-37, forewing pad 88-110, hindwing pad 11-12, caudal plate 98-101. Tarsal arolium oval (Fig. 31). Measurements as in Table 2.

Distribution: T. galii is widely distributed in the West Palaearctic and Central Asia. We have not seen any specimens from the East Palaearctic where the species probably also occurs. It has been reported from all over the Palaearctic including Japan and Taiwan (Aulmann, 1913; Klimaszewski, 1973; Gegechkori \& Loginova, 1990; Ossiannilsson, 1992) but some of the records may concern other species of the T. galii complex.

Biology: T. galii is reported to form galls on the buds, stems and leaves of Galium, Rubia and Sherardia spp. We have examined larvae and galls on following
Table 1. Measurements (in mm ) and ratios of adult Trioza species; $\mathrm{n}=$ number of measured specimens; $\mathrm{HW}=$ head width; $\mathrm{AL}=$ antenna length; WL $=$ forewing length; L3 $=$ length of antennal segment $3 ; \mathrm{WW}=$ forewing width; $\mathrm{TL}=$ metatibia length .

|  | n | HW | AL | WL | AL/HW | AL/L3 | WL/HW | WL/WW | TL/HW |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| cocquempoti | 4 0,6 우 | 0.56-0.66 | 1.18-1.31 | 2.14-2.66 | 2.03-2.28 | 3.32-4.22 | 3.45-4.56 | 2.33-2.83 | 0.90-1.11 |
| drosopouli | 5 ठ, 5 아 | 0.56-0.66 | 1.16-1.42 | 2.09-2.59 | 1.95-2.41 | 2.85-4.06 | 3.45-4.56 | 2.31-2.65 | 1.00-1.26 |
| galii | 14 ర, 20 ¢ | 0.44-0.53 | 0.69-1.04 | 1.80-2.44 | 1.38-2.29 | 2.30-4.28 | 3.75-5.20 | 2.31-4.00 | 0.87-1.33 |
| velutina | 10 す, 10 ¢ | 0.46-0.59 | 0.78-0.98 | 1.43-2.23 | 1.59-2.07 | 2.73-3.88 | 3.61-4.40 | 2.06-2.73 | 0.82-1.13 |

Table 2. Measurements (in mm) and ratios of fifth instar larva of Trioza species; $\mathrm{n}=$ number of measured specimens; $\mathrm{BL}=\mathrm{body}$ length; $\mathrm{AL}=\mathrm{an}-$ tenna length; $\mathrm{WL}=$ forewing bud length; $\mathrm{CPW}=$ caudal plate width; $\mathrm{BW}=$ body width; $\mathrm{CPL}=$ caudal plate length

|  | n | BL | AL | WL |  | CPW | BL/BW | WL/AL | BL/CPL |
| :--- | :--- | :--- | :--- | :--- | :--- | :--- | :--- | :--- | :--- |
| cocquempoti | 6 | $1.90-2.10$ | $0.40-0.45$ | $0.90-1.05$ | $1.00-1.10$ | $1.46-1.62$ | $2.22-2.75$ | $2.44-2.78$ | $0.65-0.78$ |
| drosopouli | 2 | $2.06-2.10$ | $0.40-0.45$ | $0.92-1.09$ | $1.06-1.08$ | 1.56 | $2.30-2.42$ | $2.58-2.84$ | $0.69-0.75$ |
| galii | 7 | $1.50-1.68$ | $0.30-0.32$ | $0.72-0.80$ | $0.73-0.90$ | $1.43-1.61$ | $2.25-2.67$ | $2.20-2.58$ | $0.78-0.96$ |
| velutina | - | - | - | - | - | - | - | - |  |

plants: Galium album, G. aparine, G. palustre and Asperula cynanchica. The species overwinters as adult. The adults of the new generation appear in June or July. With the available information it is impossible to deduce the number of yearly generations of $T$. galii.

Comments: The type series of Trioza galii, originally deposited in the NHMV and now in the MMBC, contains one series from Aachen and one from Ems, leg. von Heyden. The former is homogeneous, but the latter is mixed, including $1 \delta$ of T. galii and $3 \delta^{t}$ of T. velutina. As Foerster's (1848) description of T. galii is not diagnostic, and for stabilising the nomenclature, a lectotype is selected from the series from Aachen. This accords with Flor's (1861a) and Löw's (1882) concept of T. galii.

Šulc (1913) admitted that Trioza galii in his broad definition, which included $T$. distincta and T. velutina as synonyms, exhibited a surprisingly large variation in size and shape of the genal processes, in body and forewing colour, in forewing shape, venation and spinulation as well as in genital morphology. He interpreted differences in the male terminalia mostly as artefacts. For the other characters he suggested that extremes are linked by intermediates. Based on forewing characters he subdivided $T$. galii into four forms for two of which he introduced the names f. spinogalii and f . aspinovelutina, in addition to galii forma typica and f. velutina. According to paragraph 45.6 .4 of the ICZN (1999) a name introduced before 1961 as "variety" or "form" has to be considered subspecific, unless its author also expressly gave it infrasubspecific rank, or the content of the work unambiguously reveals that the name was proposed for an infrasubspecific entity, in which case it is infrasubspecific. The latter applies to the two names as Sulc mentioned that these narrowly defined forms are linked by intermediates and, thus, implying that they are artificial groupings for convenience rather than diagnosable entities such as subspecies or species. As infrasubspecific names they are not available. Boselli (1930) used the name Spanioza galii apinovelutina for Italian specimens of T. cocquempoti, thus misinterpreting Sulc's taxon, which represents T. galii judging from Šulc's figures of Austrian specimens. Boselli did not raise the name spinogalii to subspecies or species rank, and the name does not become available through Boselli (1930).

The examination of the male holotypes of T. rubiae and T. rubicunda showed that they are conspecific with T. galii, with which they are synonymised.

In Trioza chenopodii Reuter the forewing shape is variable as in T. galii. Lauterer (1982) demonstrated for the former, that this variation is seasonal, induced by day length. We found no evidence in species of the T. galii complex for seasonal differences as in T. chenopodii.

Trioza velutina Foerster, stat. rev.
Figs 4, 16-18, 26
Trioza velutina Foerster, 1848: 87. Lectotype © ${ }^{\text {T, Germany: Rheinland-Pfalz, near Boppard }}$ (A. Foerster) (MMBC), here designated, examined.

Psylla velutina; Hardy, 1853: 3876.
Trioza disticta Flor, 1861b: 401. Holotype ㅇ, Germany: Thüringen, Ruhla, mid June (G. Flor) (MMBC), examined. Syn. n.
Trioza galii sensu Šulc, 1910: 16, nec Foerster, 1848; Šulc, 1913: 45. Misidentification.
Trioza galii, Sulc, 1913: 38, p. p.
Trioza galii Foerster f. spinogalii Šulc, 1913: 44. Name unavailable, articles 45.6.1 and 45.6.4, ICZN, 1999.

Material examined: Material was examined from following countries: Algeria; Armenia; Austria: Niederösterreich, Oberösterreich, Steiermark; Croatia; Czech Republic; France: Ain, Haute-Garonne, Haut-Rhin, Nord, Pyrénées Orientales, Tarn; Germany: BadenWürttemberg, Bayern, Berlin, Brandenburg, Nordrhein-Westfalen, Rheinland-Pfalz (oै lectotype of Trioza velutina, paralectotypes 1 ठ, 2 ㅇ, Boppard), Sachsen, Thüringen (including $\circ$ holotype of Trioza distincta); Greece; Hungary; Italy: Alto Adige, Veneto; Kazakhstan; Mongolia; Russia: European part, Altay, Dagestan, Irkutsk District; Slovakia; Spain: Andalusia, Canaries; Switzerland: Aargau, Bern, Basel-Land, Basel-Stadt, Graubünden, Jura, Schaffhausen, Schwyz, Vaud; Turkey: Kars; Turkmenistan; Ukraine; United Kingdom: England, Scotland (BMNH, MHNG, MMBC, MNHN, NHMB, NHMV, SMTD, ZISP, ZMHB). The following material from the Flor collection (EAUT) was examined: 1 ठ, no 1090 on yellow label, 1 ㅇ, no 8670 on red label. Both specimens were identified by Flor as Trioza velutina. There is no list referring to these particular numbers ( O . Kurina, pers. comm.). Their provenience is therefore unknown.

Diagnosis: Adult. Total body length $2.0-2.8 \mathrm{~mm}$. Colour of thorax usually dark brown or almost black with exception of area around wing insertion which is yellowish. Younger specimens with light colour more expanded and dark areas greyish rather than black. Genal processes 0.5-0.9 times as long as vertex along mid-line, subacute apically. Forewing (Fig. 4) relatively short and broad, angular apically; bifurcation of vein M near or slightly distal to line connecting apices of veins Rs and $\mathrm{Cu}_{1 \mathrm{a}}$. Surface spinules present in all cells of forewing, forming more or less expanded fields. Hindwing three quarters of forewing length. Male proctiger (Fig. 16) relatively slender, weakly produced posteriorly. Paramere (Fig. 17) slender, lamellar, evenly tapering to apex, blunt apically. Distal portion of aedeagus (Fig. 18) with relatively short apical hook. Female proctiger (Fig. 26) with relatively short, massive apical process, pointed apically; subgenital plate, in profile, bearing small point apically. Measurements as in Table 1.

Fifth instar larva unknown.
Distribution: Probably widely distributed in the Palaearctic region. There are no reliable literature records as the species has been previously confused with T. galii.

Biology: Unknown, probably also on Galium spp. on which adults have been collected.

Comments: For the validity of the name f. spinogalii Šulc see comments on T. galii.

Trioza velutina Foerster var. thoracica Flor, nomen dubium
Trioza velutina Foerster var. thoracica Flor, 1861a: 514. Holotype ${ }^{\circ}$, France: Bouche du Rhône, Gémenos near Marseille (G. Flor), not examined, probably lost (Šulc, 1913: 44).
Comments: Flor (1861a) described Trioza velutina var. thoracica for a single $\circ$ from Southern France. According to paragraph 45.6 .4 of the ICZN (1999) the name thoracica has to be regarded as subspecific and as such becomes available. Flor's description is not diagnostic. Furthermore the holotype could not be traced in the collections of the EAUT (O. Kurina, pers. comm.) and NHMV (H. Zettel, pers. comm.) and appears to be lost (Šulc, 1913: 44). For this reason we regard thoracica as nomen dubium.

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## REFERENCES

Aulmann, G. 1913. Psyllidarum Catalogus. W. Junk, Berlin, 92 pp.
BaEva, V.G. 1972. New species of psyllids (Homoptera, Psylloidea) from Turkmenia. Doklady Akademii nauk Tadzhikistoi SSR 15: 61-63. (In Russian).
Boselli, F.B. 1930. Studii sugli psyllidi (Homoptera: Psyllidae o Chermidae) IV. Biologia e sviluppo della Spanioza galii aspinovelutina (Sulc). Bollettino del Laboratorio di Zoologia generale e agraria del R. Istituto Superiore agrario di Portici 23: 13-27.
Burckhardt, D. \& Couturier, G. 1994. The plant-louse Leuronota calycophylli sp. n. (Homoptera, Psylloidea), a pest on the timber species Calycophyllum spruceanum (Rubiaceae) in Peru. Bulletin of Entomological Research 84: 307-312.
Conci, C. 1992. Spanioza tamaninii sp. n., from Trentino (NE Italy) (Homoptera Psylloidea). Atti della Societá Italiana di Scienze Naturali e del Museo Civico di Storia Naturale di Milano 132 (1991): 257-264.
Docters van Leeuwen, W.M. 1937. New and noteworthy zoocecidia from the Netherlands. Marcellia 29: 73-86.
Edwards, J. 1896. The Hemiptera-Homoptera (Cicadina and Psyllina) of the British Isles. L. Reeve and Co., London, 271 pp .
Enderlein, G. 1926. Psyllidologica VIII. Entomologische Mitteilungen 15: 397-401.
Flor, G. 1861a. Die Rhynchoten Livlands, II. Abtheilung Sternorrhyncha Am. et S., 2. Zunft Phytophthires Burm. Dorpat, pp. 436-546.
Flor, G. 1861b. Zur Kenntnis der Rhynchoten. Bulletin de la Société Impériale des Naturalistes de Moscou 34: 331-422.
Foerster, A. 1848. Uebersicht der Gattungen und Arten in der Familie der Psylloden. Verhandlungen des naturhistorischen Vereins der preussischen Rheinlande 3: 65-98.
Gegechkori, A.M. \& Loginova, M.M. 1990. The psyllids (Homoptera, Psylloidea) of the USSR (an annotated check list). Akademiya Nauk Gruzinskoy SSR, Tbilisi, 162 pp. (In Russian).
Hardy, J. 1853. On some excrescences, etc., on plants occasioned by mites. Proceedings of the Berwickshire Naturalists' Club 3 (3): 111-113. Reprinted in Zoologist 11: 3875-3877.
Houard, C. 1909. Les zoocécidies des plantes d'Europe et du Bassin de la Méditerranée, volume 2. Librairie scientifique A. Hermann et Fils, Paris, pp. 573-1247.

ICZN International Comission on Zoological Nomenclature 1999. International Code of Zoological Nomenclature, 4th edition. International Trust for Zoological Nomenclature, London, 306 pp .
Kieffer, J.J. 1889. Mittheilungen über Psylliden. Entomologische Nachrichten 14: 222-224.
Klimaszewski, S.M. 1973. The jumping plant lice or psyllids (Homoptera Psylloidea) of the Palaearctic: an annotated check list. Annales zoologici, Warszawa 30: 155-286.
LaUterer, P. 1982. New data on the occurrence, bionomics and taxonomy of some Czechoslovakian Psylloidea (Homoptera). Acta Musei Moraviae, Scientiae naturales 67: 133-162.
Lethierry, L. 1874. Catalogue des Hémiptères du Département du Nord, 2nd edition. Imprimerie L. Danel, Lille, 108 pp., 2 plates.

Loginova, M.M. 1978. New species of psyllid (Homoptera, Psylloidea). Trudy Zoologicheskogo Instituta 61: 30-123. (In Russian).
Löw, F. 1882. Revision der paläarktischen Psylloden in Hinsicht auf Systematik und Synonymie. Verhandlungen der zoologisch-botanischen Gesellschaft Wien 32: 227-254.
Oshanin, B. 1907. Verzeichnis der palaearktischen Hemipteren mit besonderer Berücksichtigung ihrer Verteilung im Russischen Reiche, volume 2 Homoptera, part 2. Druckerei der Kaiserlichen Akademie der Wissenschaften, St. Petersburg, pp. 193-384.
Ossiannilsson, F. 1992. The Psylloidea (Homoptera) of Fennoscandia and Denmark. Fauna Entomologica Scandinavica 26, 346 pp.
Scott, J. 1876. Monograph of the British species belonging to the Hemiptera-Homoptera, family Psyllidae; together with the description of a genus which may be expected to occur in Britain. Transactions of the entomological Society 4: 525-569, plates 8-9.
Šulc, K. 1910. Monographia generis Trioza Foerster, pars I. Sitzungsberichte der königlich-böhmischen Gesellschaft für Wissenschaften II Klasse 1-34, plates 1-10.
Šulc, K. 1913. Monographia generis Trioza Foerster, pars IV. Sitzungsberichte der königlichböhmischen Gesellschaft für Wissenschaften II Klasse 1-48, plates 34-48.
Tuthill, L.D. 1943. The psyllids of America north of Mexico (Psyllidae: Homoptera). Iowa State College Journal of Science 17: 443-660.
Wagner, W. \& Franz, H. 1961. Überfamilie Sternorrhyncha (Psylloidea). In: Franz, H. (ed.). Die Nordostalpen im Spiegel ihrer Landtierwelt, vol. 2. Innsbruck, pp. 158-179.
White, I.M. \& Hodkinson, I.D. 1982. Psylloidea (Nymphal Stages) Hemiptera, Homoptera. Handbooks for the Identification of British Insects 2(5b), 50 pp .

